C.albicans Detection Kit v1

USER MANUAL

For *in vitro* Diagnostic Use

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IVD CE

Contents

1. PRODUCT DESCRIPTION

Bosphore® C.albicans Detection Kit v1 detects C.albicans DNA in human biological samples such as blood, blood culture, epithelial cells, semen, urine, prostatic secretions. The analytic sensitivity is 100 copies/ml. A region within the Internal Transcribed Spacer 2 (ITS2) gene is amplified and fluorescence detection is accomplished using the FAM filter.

An internal control has been integrated into the kit in order to check PCR inhibition. The amplification data of the internal control is detected with the HEX filter. The internal control is added in PCR step.

2. CONTENT

Bosphore® C.albicans Detection Kit v1 is composed of Real-Time PCR reagents, an internal control and a positive control.

Component	REAGENT	100 Reactions	50 Tests	25 Tests
1	dH_2O	(1000 µl)	(1000 µl)	(500 µl)
2	PCR Master Mix	(1628 µl)	(814 µl)	(407 µl)
3	Internal Control	(22 µl)	(15 µl)	(15 µl)
4	Positive Control	(160 µl)	(80 µl)	(40 µl)

3. STORAGE

Bosphore® C.albicans Detection Kit v1 PCR reagents should be stored at -20°C. Repeated thawing and freezing (>3x) should be avoided since it may reduce sensitivity. If the components are to be used in small amounts, they should be frozen in aliquots.

While preparing the PCR; the components should not be exposed to room temperature for more than 10 min, and the detection mix components should not be exposed to light and air more than necessary, vials must be kept closed except during pipetting. We recommend preparing the PCR on a cooling block and keeping the detection mixes within a closed container.

The components maintain their stability until the expiry dates on the labels, if they are stored at advised conditions.

4. REQUIRED MATERIALS AND DEVICES

- Montania® 483, Montania® 484 or Montania® 4896 Real-Time PCR Instrument (Anatolia Geneworks), or another Real-Time PCR system with FAM and HEX filters (such as iCycler, iQ5, CFX–BioRad, LightCycler 2.0, 480-Roche, 7500 Real-Time PCR System-ABI, Stratagene Mx3005P, Mx3000P-Agilent, LineGeneK, LineGene 9600-Bioer, Rotorgene 6000, Q-Qiagen)
- 0.2 ml Thin-Wall PCR tubes, plates or strips
- Magnesia® 16 Nucleic Acid Extraction System - Magnesia® Bacterial DNA Extraction Kit or Magnesia Tissue Genomic DNA Extraction Kit (Anatolia Geneworks) / Magrev® 24 Stand - Magrev® Bacterial DNA Extraction Kit or Magrev® Tissue Genomic DNA Extraction Kit (Anatolia Geneworks) / Bosphore Tissue Genomic DNA Extraction Spin Kit or Bosphore Bacterial DNA Extraction Spin Kit (Anatolia Geneworks) or other high quality DNA extraction kits and systems
- Deep freezer (-20°C)
- Desktop centrifuge with rotor for 2 ml. microcentrifuge tubes

- Calibrated adjustable micropipettes
- DNAse, RNAse, pyrogen free micropipette tips with filters
- DNAse, RNAse, pyrogen free 1.5 or 2 ml. microcentrifuge tubes
- Disposable laboratory gloves

5. IMPORTANT NOTES AND SAFETY INSTRUCTIONS

Important!:

- The product should be delivered on dry ice. Check for presence of dry ice upon arrival.
- Check for the expiry dates on the box and tube labels, upon arrival. Do not use expired products or components.
- Calibrated or verified micropipettes, DNAse, RNAse, pyrogen free micropipette tips with filters, and DNAse, RNAse, pyrogen free microcentrifuge tubes should be used.
- Before starting a test procedure, all components should be thoroughly thawed. After thawing, all components should be centrifuged briefly (spin-down for 3-5 seconds), and mixed well to ensure homogeneity prior to use.
- The kit components should be kept on ice or a cooling block until the reaction is prepared, and they should be quickly returned to -20ºC.
- PCR and nucleic acid isolation must be performed in different compartments. Samples should be stored separately to avoid contact with the kit components.
- Pathogen information should be reviewed to be aware of the health related risks.
- Clinical samples should be handled with extreme caution: Physical contact with pathogens should be avoided by; wearing lab coats and gloves, no allowance for eating or drinking within the workspace, prevention of unauthorized individuals' access to the working area.
- All the pathogenic wastes produced during the nucleic acid isolation step; including the biological samples and material contacted with them, should be discarded into medical waste and disposed safely.

6. PRODUCT USE LIMITATIONS

- All the components may exclusively be used for in vitro diagnostics.
- This product should be used in accordance with this user manual.
- This product is to be used by personnel specially trained to perform in vitro diagnostic procedures.

7. PATHOGEN

Causative Agents

Candida Albicans, the most common species of genus 'Candida', belongs to 'Saccharomycetaceae' family. İt is composed of 16mb-long (haploid) DNA genome which has varies of chromosomal arrangements resulting in its polymorphic adaptation. C. albicans may cause 3 major type of infections; oropharyngeal candidiasis, vulvovaginal (genital) candidiasis, and invasive candidiasis (candidemia). [1]

Both immunocompromised and non- immunocompromised individuals may develop skin and mucous infections due to C. albicans. It is one of the main causative agent of nosocomial infections in intensive care units. [3]

Epidemiology

Almost 30-55% of young adults may have oropharyngeal candidiasis caused by C.Albicans which is also isolated from 40-65% of normal fecal samples. C. albicans is the main reason for invasive fungal infections in immunocompromised individuals. 90% of HIV-infected patients without antiretroviral therapy may develop oropharyngeal candidiasis. [2]

Candidemia which is common Almost in 7 of every 1000 intensive care unit patients, is the one of the main agent in opportunistic bloodstream infections that is ranked as number 4 within blood culture isolates. [3]

Modes of Transmission

Its first transmission usually occurs from mother to infant during childbirth. Normally C. albicans lives symbiotically within the mucosal membranes of human (mouth, gut, vagina and skin) without causing any damage. Imbalances in the normal bacterial flora may lead overgrowth of Candida causing symptoms of disease. Transmission occurs mostly between infected patients in intensive care units of hospitals. [3], [4]

8. METHOD

Bosphore® C.albicans Detection Kit v1 is based on the Real-Time PCR method. Polymerase chain reaction is a technique that is used for amplification of a DNA region. The reaction occurs by the repeating cycles of heating and cooling. The main components of PCR are primers, dNTPs, Taq polymerase enzyme, buffer solution and template. As a brief explanation, primers are small synthetic DNA those anneal to the specific regions of the template in order to start the synthesis. dNTPs are the building blocks of the amplified products. Taq polymerase amplifies the DNA template. Buffer solution provides the pH adjustment required for the reaction and template, as referred, is the target region for synthesis.

In Real Time PCR technique, in contrast to conventional PCR, PCR product can be monitored during the reaction. Therefore Real-Time PCR obviates the need for further analysis methods like gel electrophoresis, whereby minimizing the risk of contamination. Dual labeled probes employed in the reaction in addition to the conventional PCR reagents, enable detection of the amplified target with increased sensitivity.

The assay utilizes the 5′ exonuclease activity of Taq Polymerase to cleave a dual-labeled fluorescent hybridization probe during the extension phase of PCR.

The probe is labeled at the 5′ end with a fluorescent 'reporter' molecule, and at the 3'end with another fluorescent molecule that acts as a 'quencher' for the 'reporter'. When the two fluorophores are in close proximity, and the reporter is excited by light, no reporter fluorescence can be detected. During the elongation step of PCR, Taq Polymerase encounters and cleaves the probe bound to the template. As the reporter is freed from the suppressing effect of the quencher, fluorescence signal can be detected.

The fluorescence generated by the reporter increases as the PCR product is accumulated; the point at which the signal rises above back-ground level and becomes distinguishable, is called the threshold cycle (C_T). There is a linear relationship between the log of the starting amount of a template and its threshold cycle.

Bosphore C.albicans Detection Kit v1 employs multiplex PCR, and an internal control is incorporated into the system in order to check for possible PCR inhibition. C.albicans DNA and an internal control are co-amplified in a single reaction, using sequence-specific primers. The fluorescent signal generated by the C.albicans amplification is detected by a probe labeled at the 5′ end with FAM, through the FAM channel. The fluorescent signal generated by

the internal control amplification, is detected by a second probe (labeled at the 5′ end with a different reporter molecule, HEX) through the HEX channel.

9. PROCEDURE

9.1. DNA Isolation

Magnesia® 16 Nucleic Acid Extraction System - Magnesia® Bacterial DNA Extraction Kit or Magnesia Tissue Genomic DNA Extraction Kit (Anatolia Geneworks) / Magrev® 24 Stand - Magrev® Bacterial DNA Extraction Kit or Magrev® Tissue Genomic DNA Extraction Kit (Anatolia Geneworks) / Bosphore Tissue Genomic DNA Extraction Spin Kit or Bosphore Bacterial DNA Extraction Spin Kit (Anatolia Geneworks) or other high quality DNA extraction kits and systems are used with Bosphore® C.albicans Detection Kit v1. The DNA isolation should be performed according to the manufacturers' instructions.

9.2. Kit Components

9.2.1. PCR Master Mix

PCR mix contains a highly specific and accurate HotStarTaq DNA Polymerase, the PCR buffer, the dNTP Mix, as well as the C.albicans -specific forward and reverse primers and a dual-labeled probe and internal control-specific forward and reverse primers and a dual-labeled probe.

9.2.2. Internal Control

An internal control is included in the kit to control PCR inhibition. The internal control is a synthetic DNA molecule. It is added directly into the PCR master mix to control the PCR inhibition exclusively. For this purpose, 0.2 µl of internal control should be added for each reaction into the master mix. Lack of internal control amplification in the FAM negative samples, may indicate a problem in PCR or PCR inhibition. In this case, PCR should be repeated. In samples that contain a high bacterial load, internal control can be suppressed and no increase of the signal is detected. Please use the table below for the interpretation of internal control data:

C.albicans (FAM)	Internal Control (HEX)	Interpretation
+	+	Sample positive
-	+	Sample negative
+	-	Sample positive
-	-	Repeat the test!

9.2.3. Positive Control

The positive control contains C.albicans DNA. It can be included in the PCR to test the efficiency of the PCR exclusively. The threshold cycle for the positive control is given in the acceptance criteria table (Section 10. Analysis). Threshold cycles higher than the acceptance criteria may indicate an efficiency loss in the reaction.

9.3. Preparing the PCR

Positive control should be added into the PCR reaction together with the samples and the negative control (PCR-grade water) in order to deduce the positivity/negativity of the samples. Make sure that all the kit components are thawed before use. Refer to the table below for preparing the PCR. It is for only one reaction, multiply these values with the sample number to find the values required for the master mix. While preparing master mixes for more than 5 samples, an extra 10% should be added to the total sample number.

PCR Master Mix	14.8 µl
Internal Control	0.2 µl
Sample DNA	10 µl
Negative/Positive Control	
Total Volume	25 µl

Pipette 15 µl of the master mix into the PCR tubes or strips, and add 10 µl of DNA (sample/positive or negative control). Close the tube cap. Make sure that the solution in each tube is at the bottom of the tube. Centrifuge if necessary.

9.4. Programming the Real-Time PCR Instrument

The thermal protocol for Bosphore® C.albicans Detection Kit v1 is composed of an initial denaturation for activation the HotStarTaq DNA Polymerase, a two-step amplification cycle and a terminal hold. The real-time data is collected at the second step of the amplification cycle.

Initial denaturation	95°C	14:30 min.	
Denaturation	97°C	00:30 min.	} 50 cycles
Annealing and Synthesis (Data Collection)	56°C	01:30 min.	
Hold	22°C	05:00	

Before starting a Real-Time PCR reaction using the Bosphore® Kits, the following steps should be completed:

- Choose the filter pairs to be used (FAM and HEX),
- Identify unknown samples, positive and negative controls,
- Select the correct thermal protocol,
- Start the experiment.

10. ANALYSIS

By the end of the thermal protocol, the Real-Time PCR Instrument software automatically calculates the baseline cycles and the threshold.

Example of an amplification curve is given in Fig. 1.

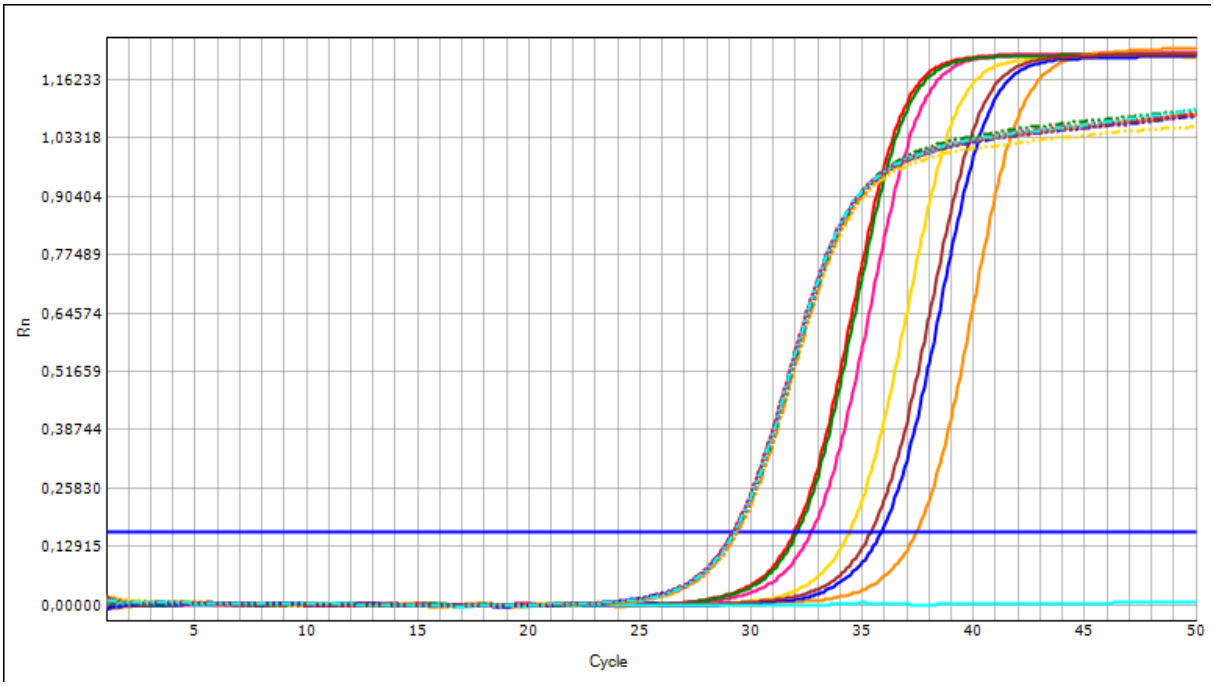

Fig. 1: Amplification Curve of a Bosphore® C.albicans v1 test

Analysis of the results should be performed by trained personnel who have received the required training for analyzing Real-Time PCR data. We recommend that the test results must be evaluated by an expert clinician, taking the patient's clinical findings and the results of other tests into consideration.

All analysis is done automatically in routine use. However, when the trained personnel, who have received the required training from manufacturer, consider it as necessary, if the system allows pulling down the threshold as much as possible in order to detect low positive samples, attention should be paid to keep the threshold line above the background and to keep the correlation coefficient at the maximum possible value (and within its acceptance criteria.)

Positive control of Bosphore® C.albicans v1 is essential for accurate result analysis. The cycle threshold acceptance criteria for the positive control is 32±4 . Test results should not be reported unless the assay results meet the criteria stated above. Please contact the manufacturer if impairment in the product's performance is observed (See the last page for contact information).

The qualitative results of the test are displayed on the "Report Mode" screen. The samples that cross the threshold in Channel 1 (FAM) are displayed as positive whereas samples that do not cut the threshold are displayed as "Negative" or "No Ct". These samples are regarded as negative or having a bacterial load below the detection limit of the assay. For these undetectable samples, the HEX data of the internal control should also be checked to avoid false negative results.

The following table shows the possible results and their interpretation:

Signal detected in FAM filter pair	The sample contains C.albicans DNA, the result is positive	No need to check the internal control since the sample is positive (high positive samples may suppress the signal from the internal control)
No signal in FAM, signal in HEX	The C.albicans DNA in the sample is not detectable	Signal from HEX filter pair rules out the possibility of PCR inhibition
No signal in FAM and HEX	The diagnosis is inconclusive	No signal in HEX points out to PCR inhibition or to a problem in DNA isolation

11. SPECIFICATIONS

11.1. Sensitivity

Analytical sensitivity may be expressed as the limit of detection: i.e. the smallest amount of the target marker that can be precisely detected. The detection limit of an individual analytical procedure is the lowest amount of nucleic acid in a sample which can be detected but not necessarily quantitated as an exact value. The analytical sensitivity or detection limit for NAT assays is expressed by the 95% positive cut-off value.

The analytical detection limit for Bosphore® C.albicans v1 was found to be 100 copies/ml. The sensitivity was determined using a dilution series of a quantitated DNA Control. The dilutions were tested in different runs in replicates. The results were analyzed by probit method.

11.2. Cross-Reactivity

To eliminate potential cross-reactivity, both assay design evidence and experimental studies were employed. Primer and probe sequences were checked for possible homology to other known pathogen sequences by sequence comparison analysis using database alignment. Samples of Aspergillus, Haemophilus influenzae, Neisseria meningitidis, Klebsiella pneumoniae, Streptococcus pneumoniae, CMV, EBV, C.trachomatis, Parvovirus B19 and MTBC with known high positivity were tested, and found negative.

12. REFERENCES

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3- Centers for Disease Control and Prevention. Candidiasis. [<http://www.cdc.gov/fungal/diseases/candidiasis/index.html/>]

4- Rangel-Frausto, M. S., Houston, A. K., Bale, M. J., Fu, C., & Wenzel, R. P. (1994). An experimental model for study of Candida survival and transmission in human volunteers. *European Journal of Clinical Microbiology and Infectious Diseases, 13*(7), 590-595.

13. SYMBOLS


Use by

Lot/Batch

Catalog number


Temperature limitation


Caution, consult accompanying documents


Manufacturer

In Vitro Diagnostic Medical Device

14. CONTACT INFORMATION

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